

# Pediatric Spinal Cord Injury: The Challenges of Injury Modelling and Translation to the Clinic

Theresa C Sutherland<sup>1\*</sup>  
Catherine A Gorrie<sup>1</sup>

<sup>1</sup>Neural Injury Research Unit, School of Life Science, University of Technology Sydney, Australia

Spinal cord injury (SCI) is a devastating condition that can arise from mechanical trauma to the spinal cord, or from a variety of non-traumatic insults, such as infection, oncogenesis, birth trauma, and electrocution [1]. Regardless of the cause, SCI will result in either complete or partial loss of motor and sensory function below the lesion site [2], as well as some degree of autonomic dysfunction [3]. SCI will often result in severe loss of tissue and varying degrees of functional impairment, and, after SCI, the spinal cord exhibits only limited repair [4]. This can have debilitating effects on the quality of life, and even the life expectancy, of SCI patients [5].

In the adult population, the majority of SCI results from motor vehicle accidents (MVA) [6]. In infants and children, the common causes of SCI include trauma, resulting from MVA and sports injury, but also from infections, neoplasms, congenital malformations, and birth trauma [7]. The majority of SCI occur at the cervical level [2], resulting in more severe autonomic dysfunction and a greater loss of function in the body than a similar injury lower in the cord. SCI has a high cost to the community, both financially and socially, although there is a lack of accurate epidemiological data available in many countries [1]. A 2007 estimate of the global incidence of spinal cord injury resulting from trauma (TSCI) was 23 cases per million population each year [1]. Less is known about pediatric SCI, as it is rarer, accounting for only 1-13% of all SCI [7-10]; however, pinning down an exact figure is difficult as different studies use different age ranges and different parameters to assess the injury based on hospital admissions, ASIA score and associated co-morbidities [7-10]. In the pediatric SCI population, the majority of injuries result from non-traumatic SCI, with traumatic spinal cord injury (TSCI) being much less common [7].

SCI has a biphasal pathophysiology consisting of the primary, immediate injury and a prolonged, exacerbating secondary injury phase [18-21]. There is little that can be done in the primary injury phase and the secondary damage phase of SCI is complex and changes over time, making it difficult to identify a simple therapeutic target to alleviate its detrimental effects. This injury phase involves multiple mechanisms and systems, not the least of which is the inflammatory response, however we still have little understanding of how these may differ between mature and pediatric patients and animal models. The inflammatory response plays a significant role in the profile of the microenvironment of the lesion after SCI, as do the actions of reactive astrocytes and activated endogenous microglia [22]. This basic pathophysiology is common to SCI in both adult and developing cords. The majority of SCI research has been carried out in animal models with a variety of different mammals used in adult models, including non-human primates. Pediatric models have used pigs [23], cats [24-26], and possums [27] as well as the common use

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**\*Corresponding author:** Theresa C Sutherland, Neural Injury Research Unit, School of Life Sciences, University of Technology Sydney, Australia. Email: [Theresa.C.Sutherland\(at\)student.uts.edu.au](mailto:Theresa.C.Sutherland(at)student.uts.edu.au)

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of mice [28-30] and rats. This has given a broad view of the similar response in a wide range of mammals, although little has been corroborated in humans. However, as mammals, it is thought that humans will exhibit a similar response to that of the experimental animals used in research [31]. The developing spinal cord exhibits significant difference to the fully developed adult cord in a variety of aspects, from biomechanical [32-34], cellular and structural [23,35,36] to molecular [28,37-39]. There is also a trend for infants having a better recovery from analogous injury than their adult counterparts, that bears greater scrutiny [14,32,35,40,41].

All of this causes some difficulty in exploring SCI in the pediatric population experimentally. Despite the prevalence of non-traumatic SCI in the pediatric population the vast majority of work exploring pediatric SCI is performed using traumatic models of injury. This is due to the complexities in creating an infant model; traumatic models are logistically easier, more readily reproducible and comparable to similar models in adults. We also have only a limited understanding of the analogous ages between the model animals and human development, as well as the developmental timing. The developmental timing, and especially the landmark development stages, are poorly understood in our model animals which creates difficulty in aligning these models with the same landmarks in human development. This alignment is necessary to account for the impact that the development of the spinal cord, CNS and exogenous systems is having on the response to a SCI in the pediatric population. To further validate these models, allow for greater utility in studying the pathophysiology of SCI and for the development of potential therapies a deeper understanding of the model animals themselves is essential.

The 'normal' behavior of infant and neonatal animals is inherently different to that in fully developed adults, and changes with different stages of development, which also adds another layer of complexity to analyzing models of pediatric SCI. In a pediatric model of SCI, it is hard to accurately ascertain where development ends and recovery begins. Very little is known about how much of an impact the developmental state and plasticity of young spinal cords has on injury recovery and the potential of 'rewiring' around the injury. This is further complicated by the presence of central pattern generation in the spinal cord. Central pattern generation allows for the development of reflex movements, without significant input from descending pathways and is common in infant animals. This complicates the assessment of locomotor function in these animals after injury.

SCI in the pediatric population may be rarer, however it is an injury that incurs literally 'life-long' ramifications. Unfortunately, we still understand very little about how the developing spinal cord responds to injury, or how the state of development affects this response. Pediatric SCI is quite a unique injury and therefore presents unique challenges on a clinical level, as well as ongoing challenges for the patient due to its great effect on ongoing physical and psycho-social development [42]. Injury presentation and aetiology of pediatric SCI is different to that in mature adults on a basic and clinical level, and a greater understanding of

the mechanisms behind SCI in younger subjects is needed to assist in the clinical management of these patients. The development of clinically relevant animal models is challenging and still requires substantial exploration. While current traumatic SCI models have found some promising avenues of research and a trend of better recovery in younger animals the developmental and behavioral complexities inherent in a pediatric model of SCI need to be addressed. And finally, a greater effort needs to be devoted to finding models to understand the progression of non-traumatic injuries as well as the post-injury sensory and autonomic impacts.

## References

1. Chang S and Hou C (2014) An Overview of Traumatic Spinal Cord Injury. In: Functional Bladder Reconstruction Following Spinal Cord Injury via Neural Approaches 1-7.
2. Barnabé-Heider F and Frisén J (2008) Stem cells for spinal cord repair. *Cell Stem Cell* 3: 16-24.
3. Karlsson A-K (2006) Overview: Autonomic dysfunction in spinal cord injury: clinical presentation of symptoms and signs. *Progress in brain research* 152: 1-8.
4. Schwartz M and Yoles E (2006) Immune-based therapy for spinal cord repair: autologous macrophages and beyond. *Journal of neurotrauma*, 23: 360-370.
5. Mathias C (2008) Autonomic dysfunction in spinal cord injury. In: *Proceedings of The Physiological Society: 2008: The Physiological Society*.
6. Norton L (2010) Spinal Cord Injury, Australia, 2007-08. Australian Institute of Health and Welfare.
7. Lee JH, Sung IY, Kang JY, Park SR (2009) Characteristics of pediatric-onset spinal cord injury. *Pediatrics International* 51: 254-257.
8. Apple DF, Anson CA, Hunter JD, Bell RB (1995) Spinal cord injury in youth. *Clinical pediatrics* 34: 90-95.
9. Karimi-Abdolrezaee S, Eftekharpour E, Wang J, Schut D, Fehlings MG (2010) Synergistic effects of transplanted adult neural stem/progenitor cells, chondroitinase, and growth factors promote functional repair and plasticity of the chronically injured spinal cord. *Journal of Neuroscience* 30: 1657-1676.
10. Osenbach RK and Menezes AH (1989) Spinal cord injury without radiographic abnormality in children. *Pediatric Neurosurgery* 15: 168-175.
11. DeVivo MJ and Vogel LC (2004) Epidemiology of Spinal Cord Injury in Children And Adolescents. *The Journal of Spinal Cord Medicine* 27: S4-S10.
12. Armstead WM (2000) Age-dependent cerebral hemodynamic effects of traumatic brain injury in newborn and juvenile pigs. *Microcirculation* 7: 225-235.
13. Dickman CA, Rekeate HL, Sonntag VK, Zabramski JM (1989) Pediatric spinal trauma: vertebral column and spinal cord injuries in children. *Pediatric Neurosurgery* 15: 237-256.
14. Parent S, Mac-Thiong J-M, Roy-Beaudry M, Sosa JF, Labelle H (2011) Spinal cord injury in the pediatric population: a systematic review of the literature. *Journal of neurotrauma* 28: 1515-1524.
15. Kunkel-Bagden E, Dai H-n, Bregman BS (1993) Methods to assess the development and recovery of locomotor function after spinal cord injury in rats. *Experimental neurology* 119: 153-164.
16. Balasingam V, Tejada-Berges T, Wright E, Bouckova R, Yong VW: Reactive astrogliosis in the neonatal mouse brain and its modulation by cytokines. *Journal of Neuroscience* 1994, 14(2):846-856.
17. Schottler J, Vogel LC, Sturm P (2012) Spinal cord injuries in young children: a review of children injured at 5 years of age and younger.

- Developmental Medicine & Child Neurology 54: 1138-1143.
18. Donnelly DJ and Popovich PG (2008) Inflammation and its role in neuroprotection, axonal regeneration and functional recovery after spinal cord injury. *Experimental neurology* 209: 378-388.
19. Kwon BK, Tetzlaff W, Grauer JN, Beiner J, Vaccaro AR (2004) Pathophysiology and pharmacologic treatment of acute spinal cord injury. *The Spine Journal* 4: 451-464.
20. Ross MH and Pawlina W (eds.) (2011) *Histology: A Text and Atlas*, Lippincott Williams & Wilkins.
21. Rowland JW, Hawryluk GW, Kwon B, Fehlings MG (2008) Current status of acute spinal cord injury pathophysiology and emerging therapies: promise on the horizon. *Neurosurgical focus* 25: E2.
22. Sutherland TC, Mathews KJ, Mao Y, Nguyen T, Gorrie CA (2017) Differences in the cellular response to acute spinal cord injury between developing and mature rats highlights the potential significance of the inflammatory response. *Frontiers in cellular neuroscience* 10: 310.
23. Kuluz J, Samdani A, Benglis D, Gonzalez-Brito M, Solano JP, et al. (2010) Pediatric spinal cord injury in infant piglets: description of a new large animal model and review of the literature. *The journal of spinal cord medicine* 33: 43.
24. Bregman BS and Goldberger ME (1983) Infant lesion effect: III. Anatomical correlates of sparing and recovery of function after spinal cord damage in newborn and adult cats. *Developmental Brain Research* 9: 137-154.
25. Bregman BS and Goldberger ME (1983) Infant lesion effect: II. Sparing and recovery of function after spinal cord damage in newborn and adult cats. *Developmental Brain Research* 9: 119-135.
26. Bregman BS and Goldberger ME (1983) Infant lesions effect: I. Development of motor behavior following neonatal spinal cord damage in cats. *Developmental Brain Research* 9: 103-117.
27. Lane M, Truettner J, Brunschwig JP, Gomez A, Bunge M, et al. (2007) Age-related differences in the local cellular and molecular responses to injury in developing spinal cord of the opossum, *Monodelphis domestica*. *European Journal of Neuroscience* 25: 1725-1742.
28. Kumamaru H, Saiwai H, Ohkawa Y, Yamada H, Iwamoto Y, et al. (2012) Age-related differences in cellular and molecular profiles of inflammatory responses after spinal cord injury. *Journal of cellular physiology* 227: 1335-1346.
29. Hamilton L, Truong M, Bednarczyk M, Aumont A, Fernandes K (2009) Cellular organization of the central canal ependymal zone, a niche of latent neural stem cells in the adult mammalian spinal cord. *Neuroscience* 164: 1044-1056.
30. Xu R, Wu C, Tao Y, Yi J, Yang Y, et al. (2008) Nestin-positive cells in the spinal cord: a potential source of neural stem cells. *International Journal of Developmental Neuroscience* 26: 813-820.
31. Norenberg MD, Smith J, Marcillo A (2004) The pathology of human spinal cord injury: defining the problems. *Journal of neurotrauma* 21: 429-440.
32. Clarke EC and Bilston LE (2008) Contrasting biomechanics and neuropathology of spinal cord injury in neonatal and adult rats following vertebral dislocation. *Journal of neurotrauma* 25: 817-832.
33. Clarke EC, Cheng S, Bilston LE (2009) The mechanical properties of neonatal rat spinal cord in vitro, and comparisons with adult. *Journal of biomechanics* 42: 1397-1402.
34. Maisonpierre PC, Belluscio L, Friedman B, Alderson RF, Wiegand SJ, et al. (1990) NT-3, BDNF, and NGF in the developing rat nervous system: parallel as well as reciprocal patterns of expression. *Neuron* 5: 501-509.
35. Vega-Avelaira D, Moss A, Fitzgerald M (2007) Age-related changes in the spinal cord microglial and astrocytic response profile to nerve injury. *Brain, behavior, and immunity* 21: 617-623.
36. Firkins SS, Bates CA, Stelzner DJ (1993) Corticospinal tract plasticity and astroglial reactivity after cervical spinal injury in the postnatal rat. *Experimental neurology* 120: 1-15.
37. Bregman BS, McAtee M, Dai HN, Kuhn PL (1997) Neurotrophic factors increase axonal growth after spinal cord injury and transplantation in the adult rat. *Experimental neurology* 148: 475-494.
38. Nakamura M and Bregman BS (2001) Differences in neurotrophic factor gene expression profiles between neonate and adult rat spinal cord after injury. *Experimental neurology* 169: 407-415.
39. Blesch A and Tuszynski MH (2009) Spinal cord injury: plasticity, regeneration and the challenge of translational drug development. *Trends in neurosciences* 32: 41-47.
40. Brown KM, Wolfe BB, Wrathall JR (2005) Rapid functional recovery after spinal cord injury in young rats. *Journal of neurotrauma* 22: 559-574.
41. Kunkel-Bagden E, Dai H-n, Bregman BS (1992) Recovery of function after spinal cord hemisection in newborn and adult rats: differential effects on reflex and locomotor function. *Experimental neurology* 116: 40-51.
42. Augutis M (2007) Pediatric spinal cord injury. *Karolinska Universitetssjukhuset*.